

## pRPa<sup>iTAG</sup>

For inducible expression of N/C-terminal human cMYC- (EQKLISEEDL) or N/C-terminal enhanced GFP-tagged proteins in *T. brucei* from a tetracycline-responsive RRNA promoter.

- High fidelity polymerase recommended.
- Integrates at the tagged RRNA spacer in 2T1/TAG<sup>PAC</sup> *T. brucei* following digestion with *AscI* (Alsford et al, 2005), giving a transformation efficiency of  $\sim 2.5 \times 10^{-6}$  (Alsford & Horn, 2007).

### Cloning GFP/MYC<sub>X</sub>

To ensure that your gene is in frame with the tag, place the second codon downstream of the *XbaI* site, i.e. TCTAGA:[codon 2]:[codon]<sub>n</sub>:[stop]:GGATCC.

### X<sup>GFP</sup>

To ensure that your gene is in frame with the tag, place the last but one codon upstream of the *XbaI* site, i.e. AAGCTT:[start]:[codon]<sub>n-1</sub>:TCTAGA.

### X<sup>6MYC</sup>X

**N-terminal tagging:** clone your gene without a start codon via *AvrII*/*Bam*HI digestion (or without start/stop codons via *AvrII*)  
i.e. CCTAGG[codon 2]:[codon]<sub>n</sub>:[stop]:GGATCC.

**C-terminal tagging:** clone your gene without a stop codon via *Hind*III(or *PacI*)/*XbaI* digestion (without start/stop codons via *XbaI*)  
i.e. AAGCTT:[start]:[codon]<sub>n</sub>:TCTAGA.

Use *Hind*III / *Bam*HI if you don't want the tag.

There are alternatives if the gene contains *XbaI*, *AvrII* and/or *Bam*HI:

Plasmid	<i>XbaI</i> , <i>AvrII</i> <i>Bam</i> HI	Insert	<i>AvrII</i> , <i>NheI</i> , <i>SpeI</i> , <i>XbaI</i> <i>Bgl</i> II
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### Key features

- Complete sequences available.
- Hygromycin for stable selection.
- All vectors allow inducible expression using tetracycline (or analogues).
- Inducible cassette is independent of selectable marker.
- Modular nature allows tag or other components to be exchanged.
- Compatible with *T. brucei* cell lines expressing TetR and containing a tagged RRNA spacer, e.g. 2T1/TAG<sup>PAC</sup> (Alsford et al, 2005).

Upon integration into *T. brucei*, the construct replaces TAG<sup>PAC</sup> and generates a functional HYG<sup>R</sup> at the previously tagged RRNA spacer. The operator binds Tet-repressor in the absence of tetracycline so the inducible RRNA promoter is activated and tagged protein is expressed when tetracycline (1  $\mu$ g ml<sup>-1</sup>) is added to the medium.

### Detection:

cMYC      Mouse anti-cMYC, 9E-10 (Source Biosciences; IFA / western blotting)  
            Mouse anti-cMYC, 4A6 (Upstate Biotechnology; western blotting only, as binds the mitotic spindle in *T. brucei*)

eGFP      Rabbit anti-GFP, IgG fraction (Molecular Probes; IFA and western blotting)

Other questions/comments, contact Sam Alsford ([sam.alsford@lshtm.ac.uk](mailto:sam.alsford@lshtm.ac.uk)).

## pRPa<sup>iSL</sup>

For inducible expression of stem loop RNA in *T. brucei* from a tetracycline-responsive RRNA promoter.

- Primers: We use a software tool (RNAit – Redmond et al, 2003) for the selection of RNAi targets that provides primer information and minimises off-target effects (see Durand-Dubief et al, 2003).
- Integrates at the tagged RRNA spacer in 2T1/TAG<sup>PAC</sup> *T. brucei* after *AscI* digestion (Alsford et al 2005), giving a transformation efficiency of  $\sim 2.5 \times 10^{-6}$  (Alsford & Horn, 2007).

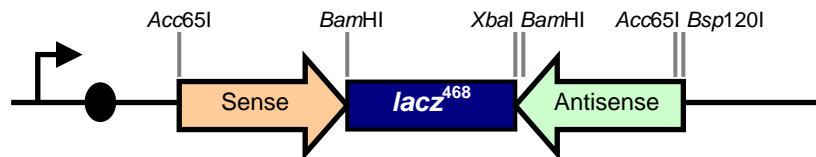
### Primer Design & Cloning

A single primer pair is required to generate the two inserts. Each primer contains two restriction sites; the internal sites allow cloning in a sense orientation into MCS1 or 3, while the external sites allow cloning in the antisense orientation into MCS2 or 4.

For example (cloning into pRPa<sup>SLi</sup>MCS<sup>1/2</sup>):

Primer A: 5'-GATC GGGCCC GGTACC -- target specific 5' sequence (20 bases) --  
*Bsp120I*      *Acc65I*

Primer B: 5'-GATC TCTAGA GGATCC -- target specific 3' sequence (20 bases) --  
*XbaI*      *BamHI*



To confirm the organisation of the stem loop cassette, use the sense fragment cloning restriction enzymes. In the above example, *Acc65I* will excise both fragments and *lacZ*<sup>468</sup>, while *BamHI* will only release *lacZ*<sup>468</sup>.

We regularly use two sequencing primers to confirm correct insertion:

*lacZ*<sup>468</sup> pos 50 towards sense RNAi  
*lacZ*<sup>468</sup> pos 420 towards antisense RNAi

Seq1 5'-AATAGTGGACTCTTGTTC  
Seq2 5'-AAAGGGGATGTGCTGCAAG

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